

**SOP: SP040b**

**IL-2 Capture ELISA Assay**

**Materials and Reagents:**

1. 96 well ELISA plate
2. Multi-channel pipettor
3. 1-200 $\mu$ l pipet tips
4. TBST (note 1)
5. 1% BSA-TBST
6. 0.1% BSA-TBST
7. Detection antibody: Biotinylated rat anti-mouse IL-2 monoclonal (BD PharMingen #18172D)
8. Streptavidin-HRP antibody (Zymed)
9. Capture antibody: purified rat anti-mouse IL-2 monoclonal (BD PharMingen #554424)
10. TMB+ Substrate-Chromogen developer (Dako #S1599)
11. Supernatants from T cell clones
12. IL-2 stock solution
13. 4°C cold room or fridge
14. Plate sealer

**Protocol:**

1. \_\_\_\_\_ Coat 96 well ELISA plate with 100 $\mu$ l capture antibody at a concentration of 2 $\mu$ g/ml and cover with a plate sealer (note 2).
2. \_\_\_\_\_ Incubate ELISA plate overnight at 4°C.
3. \_\_\_\_\_ Discard the capture antibody in the sink.
4. \_\_\_\_\_ Block ELISA plate with 200 $\mu$ l per well of blocking solution for 1 hour.
5. \_\_\_\_\_ Discard blocking solution into sink.
6. \_\_\_\_\_ Transfer T cell culture supernatants, the IL-2 standards, and the BSA negative control to the ELISA plate, 100 $\mu$ l/ well (note 3).
7. \_\_\_\_\_ Incubate for 1½ -2 hours at room temperature.
8. \_\_\_\_\_ Discard the supernatants into sink.
9. \_\_\_\_\_ Wash the plates with 100-200 $\mu$ l of TBST five times and on the fifth wash let stand for ten minutes.
10. \_\_\_\_\_ Prepare detection biotinylated antibody at a concentration of 1 $\mu$ g/ml in 0.1% BSA in TBST.
11. \_\_\_\_\_ Plate 100 $\mu$ l of the detection antibody and incubate for 1½ -2 hours.
12. \_\_\_\_\_ Discard secondary in sink.
13. \_\_\_\_\_ Wash the plate with TBST five times and on the fifth wash let stand for ten minutes.
14. \_\_\_\_\_ Prepare the streptavidin-HRP antibody at a dilution of 1:2500 in 0.1% BSA-TBST, add 100 $\mu$ l per well.
15. \_\_\_\_\_ Incubate at room temperature for 1 hour.
16. \_\_\_\_\_ Bring 10ml of TMB substrate to room temperature per ELISA plate.

17. \_\_\_\_\_ Discard the antibody in the sink.
18. \_\_\_\_\_ Wash the plate with TBST five times and on the fifth wash let stand for ten minutes.
19. \_\_\_\_\_ Add 100 $\mu$ l of developer to each well and watch for color change.
20. \_\_\_\_\_ After development, stop the reaction with 100  $\mu$ l of .18M H<sub>2</sub>SO<sub>4</sub>.
21. \_\_\_\_\_ Read at 450nm on a micro-plate reader.
22. \_\_\_\_\_ Allow the developer to dry in a chemical hood before discarding the ELISA plate.

**Notes:**

1. TBST is prepared with 1.21g Tris, 8.77 g NaCl, pH 7.4, 2.5 ml 20% Tween 80 or 0.5 ml Tween 80, QS to 1L with ddH<sub>2</sub>O.
2. Dilute the antibody in .1M sodium phosphate buffer pH 9.0. Make this by adding about 2.2 ml of .1M monobasic sodium phosphate to 500 ml of 0.1M dibasic sodium phosphate buffer. The pH may need to be adjusted a little. Do this by adding more monobasic until pH 9.0 is achieved. Be sure to add 36 wells for a IL-2 standards (positive control) and 1 well for a BSA negative control
3. The stock solution of IL-2 is at 200ng/ml. From this solution, make a standard stock of 20ng/100 $\mu$ l. Take 200 $\mu$ l of this and transfer it to the ELISA well designated for 20ng/100 $\mu$ l. From this, make 2 fold dilutions (diluting in 0.1% BSA-TBST) all the way down to .01ng/ $\mu$ l. It is best to do a set of three standards to make a nice standard curve.