SOP: AB106.1

Modified: 7/14/22 KE

Isotyping of Hybridoma Monoclonal Antibodies

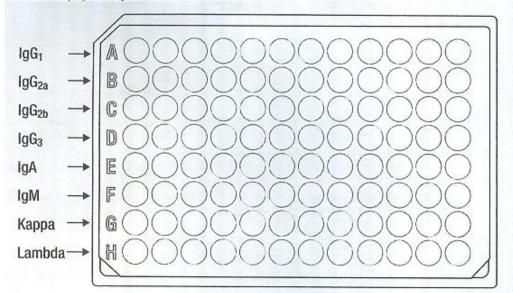
Materials and Reagents:

- 1. Pierce Rapid ELISA Mouse mAb Isotyping kit (ThermoFisher. Invitrogen Cat# 37503)
 - Protocol based off kit manual (see references for link)
- 2. TBS
- 3. Multichannel pipet with 200 µL pipet tips (use filter tips if handling biological/biohazardous fluids)
- 4. Reagent reservoirs
- 5. 0.65 mL Eppendorf tubes
- 6. Plate sealers (clear, sterile OR non-sterile, NOT PCR sealers)
- 7. Titer plate shaker (Lab-Line Instruments, Inc., Model #4625)
 - ALL shaking steps should be performed at 200 rpm (speed "2" on titer plate shaker)
- 8. Microplate reader
 - Plate reader should be able to read absorbance

Protocol	:
1	Obtain a small aliquot of hybridoma supernatant or purified antibody.
2	Prepare the antibody to be tested by diluting in TBS as follows:
	• Hybridoma supernatants: dilute sample 1:50 in TBS: add 20 μ L of hybridoma supernatant to 980 uL of TBS. The suggested assay dilution range is 1:10-1:100.
	\bullet Purified antibody: dilute sample to 250 ng/mL in TBS. The suggested assay concentration range is 25 ng/mL-2 $\mu g/mL$
3	Equilibrate TMB substrate and plate strips (from kit) to room temperature.
4	Add 50 μL of diluted antibody sample to each well of the 8-well strip.
5	Add 50 μ L of the Goat Anti-Mouse IgG+IgA+IgM HRP Conjugate (from kit) to each well of the 8-well strip. Mix by gently tapping the plate or by gentle mixing on a plate shaker.
6	Cover plate and incubate at room temperature for 1 hr.
7	Decant plate contents (Note 1).
8	Wash plate thrice with 200 μ L/well of wash buffer (from kit) (Note 2).
9	Add 75 μ L of TMB Substrate (from kit) to each well. A blue positive response may be visible after 1 min. Signal development and intensity varies depending on antibody concentration and isotype.
10	After 5-15 minutes add 75 μ L of Stop Solution (from kit). The Stop Solution changes the color from blue to yellow.
11	Read at 450 nm on a microplate reader.
12	Interpret results using the location map below (Note 3). Each sample should have a positive response in one of the rows A-F (heavy-chain identification) and a positive response in either row G or H (light-chain identification) (Note 4).

Notes:

- 1. If plate contents contain biological/biohazardous waste, the liquid must be handled and disposed of properly as biohazardous waste (cannot be decanted into sink). However, if plate contents do NOT contain any biological/biohazardous waste, then the liquid can be decanted directly into sink. Make sure to check wells are void of residual liquid (can tap microplate a couple more times). Make sure to tap on dry spot of towel after each wash as to not contaminate microplate with backsplash of liquid on towel.
- 2. Washes should be performed *quickly*. Wash buffer should be added using a multichannel pipet (with 200 μL capacity), gently tapped, then decanted into sink (3-4 shakes). *IF washes are performed after an incubation with biological fluids/biohazardous waste, the washes must be handled and disposed of <i>properly as biohazardous waste (cannot be decanted into sink)*. The microplate should be tapped 3-4 times on a towel for liquid removal. Make sure to check wells are void of residual liquid (can tap microplate a couple more times). Make sure to tap on *dry spot* of towel after each wash as to not contaminate microplate with backsplash of liquid on towel.
- 3. Location Map (plate layout)



4. Wells with the highest response (darkest color) indicate isotype and light chain composition; lightly colored wells indicate contaminating host or myeloma antibodies.

References:

1. Pierce Rapid ELISA Mouse mAb Isotyping Kit Manual: https://assets.thermofisher.com/TFS-Assets%2FLSG%2Fmanuals%2FMAN0014495 37503 Ms mAb Isotyping Pierce RapidELISA UG.pdf