Preparation of electrocompetent M. smegmatis

Materials	and	Reagents
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- 1. M. smegmatis, 1 ml frozen stock or growing culture
- 2. Biosafety cabinet (note 1)
- 3. 51 ml sterile LB media
- 4. 10% sterile glycerol
- 5. 20% sterile tween-80
- 6. Sterile aerosol resistant pipette tips, 200µl
- 7. Pipetman, 200µl
- 8. Disposable inoculating loops
- 9. 37°C shaking incubator
- 10. Spectrophotometer, visible light
- 11. Spectrophotometer cuvettes
- 12. Sterile 1.7 ml eppendorf tubes
- 13. Cryostorage box
- 14. Freezer, -80°C
- 15. Biohazard bags
- 16. Autoclave tape
- 17. Autoclave
- 18. 50 ml conical Falcon tubes
- 19. Disposable plastic serological pipettes, 50 ml and 5 ml
- 20. Pipette aid
- 21. Allegra 6 R centrifuge
- 22. Sterile 250 ml Erlenmeyer flask
- 23. Ice bucket and ice

1	Grow a fresh 50 ml culture of <i>M. smegmatis</i> MC^2 155 in either Middlebrook 7H9-ADS-Tween or LB-Tween (0.05%) to mid-log phase ($OD^{600} = 0.5$ to 1.0).
2	Incubate cells on ice for at least 10 minutes but no longer than two hours. (note 2)
3	_ Transfer cells to chilled 50 ml conical tube.
4	Spin down cells in Allegra 6 R at 3000 rpm at 4°C for 10 minutes. Decant supernatant from pellet into autoclavable container.
5	Resuspend pellet in 40 ml cold sterile 10% glycerol.
6	Centrifuge at 3000 rpm at 4°C for 10 minutes in Allegra 6R tabletop centrifuge. Decant supernatant into autoclavable container.
7	Resuspend pellet in 40 ml cold sterile 10% glycerol.
8	Centrifuge at 3000rpm at 4°C for 10 minutes. Decant supernatant into autoclavable container.
9	Resuspend pellet in 5 ml ice cold 10% glycerol and store on ice.
10	Distribute cells in 400μl aliquots into 1.7 ml eppendof tubes and store at -70°C for up to one year.
13	Autoclave and dispose of all liquid waste generated.

Notes:

- All work should be performed in a Biosafety cabinet in order to prevent contamination. See SOP SP041 for use of the biosafety hood.
 For the following steps, keep cells as close to 0°C (on ice) as possible.